INVESTIGATION of SEVERAL AEROSOL INSECTICIDE COMPOSITIONS CONTAINING SYNERGISTS

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Abstract Two aerosol insecticide compositions were investigated in laboratory conditions against a complex of synanthropic insects. Both compositions have high acute insecticidal activity and have a long residual effect on glass, but not on plywood. It is shown that the knockdown effect was reversible on plywood on the 3^{d} and 7^{th} days after treatment for cockroaches and on the 7^{th} day for bugs. Fifteen aerosol compositions containing cypermethrin, imiprothrin alone or mixed with the synergists piperonyl butoxide and/or MGK-264 in different concentrations were prepared specially for laboratory use and were investigated. It is shown that the synergism ratio was higher in houseflies than in cockroaches in compositions containing PBO, but it was higher in cockroaches when it contained an MGK-264-like synergist.

Key Words Cockroach housefly piperonyl butoxide MGK-264 resistance

INTRODUCTION

Detoxication of many xenobiotics, which are complex ethers of organic acids, is catalyzed by various enzymatic systems, specifically monooxygenases (MO) and nonspecific esterases (NE). This is also true for synthetic pyrethroids. Representatives of two orders of the class Insecta: German cockroach and house fly, served as biological models with different levels of MO and NE activity.

Piperonyl butoxide (PBO) and MGK-264 are known as inhibitors of insect detoxication systems. Both synergists are used in various insecticide compositions, mainly in aerosol cans. The best synergism occurred in a mixture of PBO or MGK-264 with pyrethrins. Insecticide resistance can be a serious problem in house fly and cockroach control. When a resistance mechanism is metabolic, it is often possible to negate that mechanism by use of synergists (Cochran, 1987, 1994; Yu and Hsu, 1993; Wu et al., 1998). Several registered insecticide compositions include the synergists PBO and/or MGK-264, which are intended for medical disinfection in public health for control of synanthropic insects in Russia: microencapsulated "Effective Ultra" (Netherland), aerosol cans "Dikhlophos VP", "Dikhlophos Super", "Dikhlophos L", "Knockout LN" (Russia) and others.

MATERIALS and METHODS

The sensitive standard strain of German cockroach *Blattella germanica* (L.), oriental cockroach *Blatta orientalis* L., bed bug *Cimex lectularius* L., rat flea *Xenopsylla cheopis* Roths., and house fly *Musca domestica* L. (strain Cooper) have been used in this study.

The biological efficacy, i.e., direct spray, and the residual efficacy on porous (plywood) and non-porous (glass) surfaces of the aerosol formulations #1 and #2 against the insects mentioned above have been tested under laboratory conditions (temperature at $25\pm1^{\circ}$ C, humidity between $55\pm10^{\circ}$ RH).

Direct spray test: crawling insects are held in 0.5 l glass vessels (3 replicates, 20 insects each), and the aerosol can is held in a clamp and aligned so that spray is directed into the open top

of the vessels from a distance of 45 cm (dose 20 g/m²). After spraying, insects are transferred to clean containers for observation of knockdown and mortality.

Residual test: plywood and glass surfaces (200 cm², 3 replicates each) sprayed from a distance of 45 cm (dose 20 g/m²). After 24 h test insects were placed on surfaces for 15 min. Then they were transferred to clean containers for observation of knockdown and mortality.

Direct spray for flying insects: 300 flies were held in a 2 m³ glass box, and a 1 g/m³ aerosol composition was sprayed into it. Knockdown times were observed at 1% to 99% flies (min), then we calculated C_{15} (concentration of active ingredient (AI) in the air which caused 99% fly mortality at 15 min, mg/m³), Q_{15} (quantity of aerosol which caused 99% fly mortality at 15 min, mg/m³), and KT_{50} min, value has been determined graphically.

The toxicological method implies the topical treatment of the insects, which were first anesthetized with diethyl ether for 20 s. Acetone solutions (1 µl) at various concentrations were applied to the mesonotum of the house flies imago and to the prothorax of cockroaches. The level of mortality was estimated within 24-48 h. The degree of toxicity was estimated through determination of LD_{50} µg/g (the dose leading to the death of 50% of the experimental individuals). In order to determine LD_{50} , a series of logarithmically decreasing concentrations inducing 20-80% mortality of insects was prepared. Working solutions of the studied compounds (or mixtures) were obtained by successive dilution. Preparation of dilutions of the aerosol mixture from cans was in two steps: the first to remove propellant, the second to prepare a series of acetone concentrations. The data on mortality of insects as a function of the concentration of the studied compounds (or mixtures) were subject to regression analysis, with determination of LD_{50} and confidence intervals (Popov, 1965a). In studies of the joint action of compounds in mixtures, the synergism ratio (SR) was calculated according to the additive rule (Popov, 1965b). For determination of knockdown time, we fixed the time after treatment with insecticide, when the insect can no longer retain itself on the upright-drilling glass surface.

RESULTS and DISCUSSION

Investigated aerosol cans contained a mix of pyrethroids (#1 permethrin and cypermethrin, #2 imiprothrin and cypermethrin) and the synergist MGK-264. Used as a direct-acting spray, both compositions kill cockroaches and other crawling insects (Table 1). The knockdown effect was rapid in composition #2, perhaps because imiprothrin was available there. The residual effect on the glass surfaces was >21 days for all insects. On the plywood, the residual effect was short for females of the German cockroach, especially in the composition #2. German cockroach males were more sensitive to residuals than were females (Table 2).

The knockdown effect of composition #2 residuals was also investigated for male and female German cockroaches and bugs. Exposure on glass or plywood was 15 minutes, dose 20 g/

| Aerosol composition, # | Test-insect (mixed sex) | % | knockdo 5 | own at tin 10 | ne(min 30 | <u>)</u> 60 | % Killing at 24 hours |
|---------------------------|----------------------------|-----|--------------|------------------|--------------|----------------|--------------------------|
| #1 | German cockroach | 0 | 100 | 100 | 100 | 100 | 100 |
| permethrin 0.3% | Oriental cockroach | 0 | 20 | 50 | 100 | 100 | 100 |
| cypermethrin 0.2% | Bed bug | 0 | 90 | 100 | 100 | 100 | 100 |
| MGK-264 1.0% | Rat flea | 20 | 100 | 100 | 100 | 100 | 100 |
| #2 | German cockroach | 100 | 100 | 100 | 100 | 100 | 100 |
| imiprothrin 0.08% | Oriental cockroach | 80 | 100 | 100 | 100 | 100 | 100 |
| cypermethrin 0.1% | Bed bugs | 100 | 100 | 100 | 100 | 100 | 100 |
| MGK-264 0.8% | Rat flea | 100 | 100 | 100 | 100 | 100 | 100 |

Table 1. Direct spray of aerosol compositions

| Aerosol | | | % Killing at time(days) | | | | | |
|-------------------|-------------|-----|-------------------------|-----|-----|-----|-----|-----|
| composition, # | Test-insect | Sex | Surface | 1 | 3 | 7 | 14 | 21 |
| #1 | German | М | Glass | 100 | 100 | 100 | 100 | 100 |
| permethrin0.3% | cockroach | F | | 100 | 100 | 100 | 100 | 100 |
| cypermethrin 0.2% | | Μ | Plywood | 80 | 60 | 50 | 30 | 10 |
| MGK-264 1.0% | | F | - | 30 | 30 | 10 | 0 | 0 |
| | Bed bug | MF | Glass | 100 | 100 | 100 | 100 | 100 |
| | | MF | Plywood | 100 | 100 | 100 | 100 | 100 |
| | Rat flea | MF | Glass | 100 | 100 | 100 | 100 | 100 |
| #2 | German | М | Glass | 100 | 100 | 100 | 100 | 97 |
| imiprothrin 0.08% | cockroach | F | | 100 | 100 | 100 | 97 | 87 |
| cypermethrin 0.1% | | Μ | Plywood | 30 | 12 | 3 | 10 | 5 |
| MGK-264 0.8% | | F | | 23 | 2 | 0 | 10 | 0 |
| | Bed bug | MF | Glass | 100 | 100 | 100 | 100 | 100 |
| | | MF | Plywood | 90 | 90 | 50 | 60 | 50 |
| | Rat flea | MF | Glass | 100 | 100 | 100 | 100 | 100 |

Table 2. Residual activity of aerosol compositions

Table 3. Knockdown effect of aerosol composition #2, containing cypermethrin, imiprothrin, and MGK-264

| | | | | Perc | centage kn | ockdow | n in minut | es | | | |
|--------------------|-----|------|---------------|------|--------------|------------|------------|---------------|-------|-------|-------|
| Surface/ | | 1 | 5 | 10 | 15 | 20 | 30 | 60 | 120 | 24 | 48 |
| Insect | Sex | | | | | | | | | hours | hours |
| | | | | | 1st day af | ter treati | nent | | | | |
| Plywood German | М | 0 | 0 | 0 | 0 | 7±3 | 13±5 | 23 <u>+</u> 6 | 50±10 | 23±7 | 30±10 |
| cockroach | F | 0 | 0 | 0 | 0 | 0 | 17±5 | 27±5 | 33±10 | 10±3 | 23±10 |
| Glass German | М | 20±7 | 80±10 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 |
| cockroach | F | 10±3 | 40 <u>±</u> 8 | 90±5 | 90±5 | 93±7 | 100 | 100 | 100 | 100 | 100 |
| | | | | | 3rd day af | fter treat | nent | | | | |
| Plywood | М | _ | _ | _ | 3±3 | _ | <u>8+2</u> | 28±5 | 30±10 | 12±5 | _ |
| German | | | | | | | | | | | |
| cockroach | F | - | _ | _ | 5±3 | _ | 28±10 | 27±10 | 43±15 | 2+2 | _ |
| Plywood Bed bug | MF | _ | _ | _ | 30±10 | _ | _ | 37±10 | 90±5 | 90±5 | _ |
| | | | | | 7th day af | ter treati | nent | | | | |
| Plywood German | М | _ | _ | _ | 0 | _ | 0 | 10±3 | 10±3 | 3±3 | 3±3 |
| cockroach | F | _ | _ | _ | 3±3 | _ | 10±3 | 23±5 | 20±5 | 0 | 0 |
| Glass German | М | _ | - | _ | 100 | - | 100 | 100 | 100 | 100 | 100 |
| cockroach | F | _ | _ | _ | 97 <u>+2</u> | _ | 100 | 100 | 100 | 100 | 100 |
| Plywood Bed bug | MF | - | _ | _ | 0 | _ | 13±10 | 20±5 | 70±10 | 50±15 | _ |

m², 24 h after spray (Table 3). Results show that there were slight, short, and unstable effects on insects even in the 1-7 days interval after spray.

Fifteen aerosol compositions containing cypermethrin, imiprothrin alone or mixed with the synergists piperonyl butoxide (PBO) and/or MGK-264 in different concentrations were prepared specially for laboratory use and were investigated on German cockroach males and house flies (mixed sex) with the topical method. The synergism ratio (SR) was higher in house flies than in cockroaches in compositions containing PBO, but it was higher in cockroaches when there were MGK-264 like synergists (Table 4).

| | | % a.i. in | German | | House flies | | |
|-----|---------------------|-----------|--------------------|------|--------------------|-----|--|
| | | aerosol | <u>cockroaches</u> | male | both sex | | |
| # | Composition | can | $LD_{50}, \mu g/g$ | SR | $LD_{50}, \mu g/g$ | SR | |
| 1. | Cypermethrin | 0.01 | 0.135±0.020 | _ | 0.27±0.03 | _ | |
| 2. | Cypermethrin | 0.01 | 0.054±0.016 | 2.5 | 0.19±0.06 | 1.4 | |
| 3. | MGK-264 | 0.8 | 0.087±0.023 | 1.6 | 0.16±0.07 | 1.7 | |
| 5. | Cypermethrin PBO | 0.01 | 0.087±0.025 | 1.0 | 0.10±0.07 | 1./ | |
| 4. | Cypermethrin | 0.01 | 0.096±0.028 | 1.4 | 0.13±0.02 | 2.1 | |
| 4. | PBO | 0.01 | 0.090±0.028 | 1.4 | 0.13±0.02 | 2.1 | |
| 5. | Cypermethrin | 0.01 | 0.112±0.045 | 1.2 | 0.20±0.06 | 1.4 | |
| 5. | PBO | 0.4 | 0.112±0.045 | 1.2 | 0.20±0.00 | 1.7 | |
| | MGK-264 | 0.4 | | | | | |
| 6. | Imiprothrin | 0.008 | 1.890±0.193 | - | 0.53±0.08 | | |
| 7. | Imiprothrin | 0.008 | 0.790±0.051 | 2.4 | 0.58±0.06 | 0.9 | |
| 7. | MGK-264 | 0.008 | 0.790±0.031 | 2.4 | 0.38±0.00 | 0.9 | |
| 8. | Imiprothrin | 0.008 | 1.020±0.130 | 1.9 | 0.26±0.04 | 2.0 | |
| | PBO | 0.8 | | | | | |
| 9. | Imiprothrin | 0.008 | 1.210±0.210 | 1.6 | 0.68 <u>±</u> 0.07 | 0.8 | |
| | PBO | 0.4 | | | | | |
| 10. | Imiprothrin | 0.008 | 1.890±0.663 | 1.0 | 1.05±0.12 | 0.5 | |
| | PBO | 0.4 | | | | | |
| | MGK-264 | 0.4 | | | | | |
| 11. | Cypermethrin | 0.01 | 0.210±0.041 | - | 0.65 ± 0.08 | - | |
| | Imiprothrin | 0.008 | | | | | |
| 12. | Cypermethrin | 0.01 | 0.035 ± 0.012 | 6.0 | 0.55±0.07 | 1.2 | |
| | Imiprothrin | 0.008 | | | | | |
| | MGK-264 | 0.8 | | | | | |
| 13. | Cypermethrin | 0.01 | 0.210±0.063 | 1.0 | 0.33±0.07 | 2.0 | |
| | Imiprothrin | 0.008 | | | | | |
| | PBO | 0.8 | | | | | |
| 14. | Cypermethrin | 0.01 | 0.350 ± 0.081 | 0.6 | 0.48±0.09 | 1.4 | |
| | Imiprothrin | 0.008 | | | | | |
| | PBO | 0.4 | 0.470.0.477 | ~ ~ | | • • | |
| 15. | Cypermethrin | 0.01 | 0.470±0.152 | 0.5 | 0.33±0.09 | 2.0 | |
| | Imiprothrin | 0.008 | | | | | |
| | PBO | 0.4 | | | | | |
| | MGK-264 | 0.4 | | | | | |

Table 4. Synergism ratio (SR) and toxicity of several insecticide compositions for insects

| # | Composition | % a.i. in aerosol can | Knock- down 1% (min) | Knock- down 99% (min) | KT ₅₀ (min) | C ₁₅ mg/m ³ | Q ₁₅ mg/m ³ |
|-----|---|-----------------------------|----------------------------|-----------------------------|---------------------------|--------------------------------------|--------------------------------------|
| 1. | Cypermethrin | 0.01 | 4'13" | 9'06" | 6'45" | 0.061 | 613 |
| 2. | Cypermethrin MGK-264 | 0.01 0.8 | 4'00" | 7'33" | 5'45" | 0.053 | 533 |
| 3. | Cypermethrin PBO | 0.01 0.8 | 3'26" | 6'53" | 5'15" | 0.048 | 484 |
| 4. | Cypermethrin PBO | 0.01 0.4 | 3'23" | 8'13" | 5'45" | 0.057 | 571 |
| 5. | Cypermethrin PBO MGK-264 | 0.01 0.4 0.4 | 3'18" | 7'42" | 5'30" | 0.046 | 460 |
| 6. | Imiprothrin | 0.008 | 3'13" | 22'31" | 13'00" | 0.134 | 1680 |
| 7. | Imiprothrin MGK-264 | 0.008 0.8 | 3'41" | 16'12" | 10'00" | 0.099 | 1237 |
| 8. | Imiprothrin PBO | 0.008 0.8 | 1'52" | 23'40" | 13'00" | 0.155 | 1932 |
| 9. | Imiprothrin PBO | 0.008 0.4 | 2'14" | 22'18" | 12'00" | 0.119 | 1487 |
| 10. | Imiprothrin PBO MGK-264 | 0.008 0.4 0.4 | 1'36" | 20'30" | 11'15" | 0.118 | 1469 |
| 11. | Cypermethrin Imiprothrin | 0.01 0.008 | 2'11" | 5'44" | 4'00" | 0.076 | 420 |
| 12. | Cypermethrin Imiprothrin MGK-264 | 0.01 0.008 0.8 | 2'01" | 6'27" | 4'30" | 0.087 | 483 |
| 13. | Cypermethrin Imiprothrin PBO | 0.01 0.008 0.8 | 3'25" | 11'30" | 7'30" | 0.154 | 855 |
| 14. | Cypermethrin Imiprothrin PBO | 0.01 0.008 0.4 | 2'20" | 12'23" | 7'30" | 0.159 | 885 |
| 15. | Cypermethrin Imiprothrin PBO MGK-264 | 0.01 0.008 0.4 0.4 | 2'38" | 11'41" | 7'00" | 0.174 | 967 |

Table 5. Knock-down effect, KT₅₀ and toxicity of several insecticide aerosol compositions for *Musca domestica* in glass box (1 g/m³)

When mixed with imiprothrin, the presence of MGK-264 results in lesser time for the house-fly knock-down effect and greater mortality. In a two-component mixture this effect was lower (Table 5). We know that the most fermentative activity occurred in resistant insect strains such as MO and NE. Our previous investigations show this fact to be true for house flies (Eremina and Bakanova, 1998; Vavilova et al., 1998; Eremina et al., 1999). Application of the synergist PBO with permethrin leads to an increase of susceptibility of houseflies Cooper strain at 10-fold, R-permethrin strain at 100 fold, and VP strain at 60 fold (Eremina and Roslavtseva, 1997). It was to

be expected that in house flies and cockroaches the resistant field strains synergist ratio will be higher than in sensitive laboratory strains.

REFERENCES

- Cochran, D.G. 1987. Effects of synergists on bendiocarb and pyrethrins resistance in the German cockroach (Dictyoptera: Blattellidae). J. Econ. Entomol. 80(4): 728-732.
- Cochran, D.G. 1994. Effects of 3 synergists on pyrethroid resistance in the German cockroach (Dictyoptera: Blattellidae). J. Econ. Entomol. 87(4): 879-884.
- Eremina, O.Yu., and Roslavtseva, S.A. 1997. The application of synergists for studying the activity of some enzyme systems and their involvement in insecticide action. Biol. Bull. 24(3): 251-258.
- ?remina, O.Yu., and Bakanova, E.I. 1998. Enzymes activities in resistant field populations of *Musca domestica* L. 9th IUPAC International Congress of Pesticide Chemistry. London. 2-7 August 1998. 1: 4B-039.
- Eremina, O.Yu., Bakanova, E.I., and Khrunin, A.V. 1999. Enzymes activities in resistant field populations of *Musca domestica* L. from Moscow. Proceedings of the 3rd International Conference on Urban Pests, ICUP-99, Prague, 1999. 627 pp.
- Popov, P.V. 1965a. Calculation of doses of mixtures of sciences for full additive and independent biological action of compounds. Khim. Sel'sk. Khoz. 8: 73-79.
- Popov, P.V. 1965b. Statistical analysis of experimental data using regression line "Pesticide Dose Activity". Khim.Sel'sk. Khoz. 10: 72-74.
- Vavilova, V.V., Roslavtseva, S.A., Eremina, O.Yu., and Bakanova, E.I. 1998. Monitoring of resistance to insecticides and fermentative activity at *Musca domestica* field populations. Sixth European Congress of Entomology. Ceske Budejovice, Czech Republic, August 23-29, 1998. 2: 689-690.
- Wu, D.X., Scharf, M.E., Neal, J.J., Suiter, D.R., and Bennett, G.W. 1998. Mechanisms of fenvalerate resistance in the German cockroach, *Blattella germanica* (L.). Pestic. Biochem. Physiol. 61(1): 53-62.
- Yu, S.J., and Hsu, E.L. 1993. Induction of detoxification enzymes in phytophagous insects -- roles of insecticide synergists, larval age, and species. Arch. Insect Biochem. Physiol. 24(1): 21-32.